

## 12-EPITEUPOLIN II, A NEO-CLERODANE DITERPENOID FROM *TEUCRIUM LAMIIFOLIUM*

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**Key Word Index**—*Teucrimum lamiifolium*; Labiatae; diterpenoids; neo-clerodane derivatives; 12-epiteupolin II; teuscordinon; teuflin; montanin C; 19-acetylgnaphalin.

**Abstract**—From the aerial parts of *Teucrimum lamiifolium*; a new neo-clerodane diterpenoid, 12-epiteupolin II, has been isolated, together with the previously known diterpenes teuscordinon, teuflin, montanin C and 19-acetylgnaphalin.

The structure of 12-epiteupolin II (6 $\alpha$ -acetoxy-19-hydroxy-4 $\alpha$ , 18:15, 16-diepoxy-neo-clerodane-13 (16),14-dien-20,12R-olide) was established by chemical and spectroscopic means and by correlation with montanin C.

### INTRODUCTION

In continuation of our studies on the diterpenes from *Teucrimum* species [1-9] we have now investigated *T. lamiifolium* D'Urv., a species which grows in southeastern Bulgaria. From the aerial parts of this plant we have isolated four previously known neo-clerodane diterpenoids, teuscordinon [10], teuflin [11], montanin C [8], 19-acetylgnaphalin [12], and a new diterpenoid, 12-epiteupolin II (1), whose structure and absolute configuration have been established on the basis of spectroscopic evidence, chemical transformations and correlation with montanin C (3).

### RESULTS AND DISCUSSION

12-Epiteupolin II (1),  $C_{22}H_{28}O_7$ , had an IR spectrum which was consistent with the presence of a hydroxyl group ( $3630\text{ cm}^{-1}$ ), a furan ring ( $3140, 1505, 1600$  and  $875\text{ cm}^{-1}$ ), an ester group ( $1740, 1240\text{ cm}^{-1}$ ) and a  $\gamma$ -lactone ( $1760\text{ cm}^{-1}$ ). The  $^1\text{H}$  NMR spectrum of the new diterpenoid (1, Table 1) revealed the existence of a secondary methyl group ( $\delta 1.12, d, J = 6.5\text{ Hz}$ ), a  $\beta$ -substituted furan ring (two  $\alpha$ -furan protons at  $\delta 7.45$  and one  $\beta$ -furan proton at  $\delta 6.37$ ), an  $\alpha,\alpha$ -disubstituted oxirane ring (two protons forming an AB system at 2.26 and 2.90  $J = 4\text{ Hz}$ ), and an acetoxyl group which must be placed at the C-6 position one proton doublet at  $\delta 4.80, J_1 = 15, J_2 = 4\text{ Hz}$ ). The presence of a primary hydroxyl group in 1 was revealed by the  $^1\text{H}$  NMR spectrum of its derivative obtained by treatment with chloroacetyl isocyanate. The resonance of the corresponding NH proton appeared at  $\delta 8.43$  as a singlet, while the triplet at  $4.06 (J = 12\text{ Hz})$  (H-19A, OH) was now a doublet at  $4.42 (J = 13\text{ Hz})$  and the doublet of H-19B at  $4.76$  was paramagnetically shifted at  $5.65 (J = 13\text{ Hz})$ , thus confirming that the free alcohol group of 1 was a primary one. Moreover, the  $^1\text{H}$  NMR spectrum of compound 1 (Table 1) was almost identical with that of teupolin II (2) a diterpenoid previously isolated from *T. polium* [2, 5]. In fact, the difference between the  $^1\text{H}$  NMR spectra of 1 and 2 was only in the

chemical shift of the C-17 methyl protons, which appeared at a slightly lower field in 1 ( $\delta 1.12$ ) than in 2 (0.99) (Table 1). On the other hand, comparison between the  $^{13}\text{C}$  NMR spectra of 12-epiteupolin II (1) and teupolin II (2) (Table 2) showed a significant difference in the C-8 ( $\Delta\delta + 2.8$ ) and C-10 ( $\Delta\delta - 2.1$ ) chemical shifts.

These data clearly established that the structural difference between compounds 1 and 2 must be only the configuration of its C-12 centre [13, 14]. Moreover, the (12R)-configuration of 1 was also in agreement with NOE experiments, since irradiation of the Me-17 protons of 12-epiteupolin II caused a 7% NOE enhancement of the H-12 signal ( $\delta 5.40$ ) [13, 15, 16].

Final proof that 12-epiteupolin II has the structure and absolute configuration depicted in formula 1 was obtained by acetic anhydride-pyridine treatment of compound 1. This gave a diacetyl derivative (3) identical in all respects [ $\alpha_D$ ,  $^1\text{H}$  NMR,  $^{13}\text{C}$  NMR and mmp] with natural montanin C [8, 13].

12-Epiteupolin II, montanin C [8, 13] and montanin G [16, 17] are the only neo-clerodane-20, 12-olides isolated from Bulgarian *Teucrimum* so far possessing the infrequent (12R)-configuration.

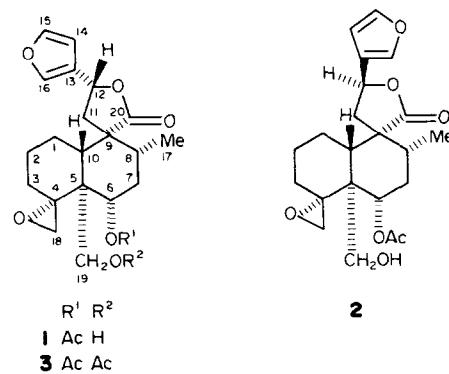


Table 1.  $^1\text{H}$  NMR data of compounds **1** and **2** (250 MHz,  $\text{CDCl}_3$  solution, TMS as int. standard)\*

H	<b>1</b>	<b>2</b>	$\Delta\text{ppm}$
$3\alpha$	†	2.50 <i>m</i>	—
$3\beta$	‡	1.09 <i>m</i>	—
$6\beta$	4.80 <i>dd</i>	4.86 <i>dd</i>	—0.06
$7\beta$	1.59 <i>m</i>	1.61 <i>m</i>	—0.02
$8\beta$	‡	2.00 <i>m</i>	—
11A	2.35 <i>d</i>	2.38 <i>d</i> (8.2 Hz)	—
11B	2.15 <i>d</i>		—
12	5.40 <i>t</i>	5.35 <i>t</i>	—0.05
14	6.37 <i>m</i>	6.39 <i>m</i>	—0.02
15	7.45 <i>m</i>	7.44 <i>m</i>	+0.01
16			
17	1.12 <i>d</i>	0.99 <i>d</i>	+0.13
18A	2.90 <i>dd</i>	2.92 <i>dd</i>	—0.02
18B	2.26 <i>d</i>	2.23 <i>d</i>	+0.03
19A	4.06 <i>t</i> ‡	4.00 <i>t</i> ‡	+0.06
19B	4.76 <i>d</i>	4.71 <i>d</i>	+0.5
COMe	2.12 <i>s</i>	2.045 <i>s</i>	+0.08

*J*(Hz) **1,2**: 6 $\beta$ ,7 $\beta$  = 4; 6 $\beta$ ,7 $\alpha$  = 15; 11A,11B = 14; 11A, 12 = 11B, 12 = 8; 17, 8 $\beta$  = 6.5; 18A,18B = 4; 18 $\Delta$ ,3 $\alpha$  = 2; 19A,19B = 13; 19A,OH = 12.

\*All these assignments have been confirmed by double resonance experiments.

†Overlapped signal.

‡Collapsed into a *d* after addition of  $\text{D}_2\text{O}$ .

## EXPERIMENTAL

Mps are uncorr.  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR spectra were obtained in  $\text{CDCl}_3$  soln with TMS as int. standard. Plant materials were collected in July 1984 near Malko Tirnovo, Bulgaria.

*Extraction and isolation of the diterpenoids.* Dried and finely powdered *T. lamiifolium* aerial parts (1.28 kg) were extracted with  $\text{Me}_2\text{CO}$  (30 l) at room temp. for a week. After evapn of the solvent, the residue was treated as in refs [18, 19]. The  $\text{CHCl}_3$  extract (29 g) was chromatographed over a silica gel column (Merck, 7734, deactivated with 10%  $\text{H}_2\text{O}$ , 500 g) eluted with petrol,  $\text{CH}_2\text{Cl}_2$ -petrol mixtures and pure  $\text{CH}_2\text{Cl}_2$ . Elution with  $\text{CH}_2\text{Cl}_2$ -petrol (2:8) gave teuflin (60 mg) and teuscordinon (40 mg); elution with  $\text{CH}_2\text{Cl}_2$  gave 19-acetylgnaphalin (38 mg), montanin C (400 mg) and 12-epiteupolin II (**1**) (80 mg).

The previously known diterpenoids, teuflin, teuscordinon, 19-acetylgnaphalin and montanin C, were identified by their physical (mp,  $[\alpha]_D$ ) and spectroscopic (IR,  $^1\text{H}$  NMR, MS) data and by comparison with authentic samples (mmp, TLC).

*12-Epiteupolin II (**1**)* Mp 173–175° (from  $\text{Me}_2\text{CO}$ -petrol),  $[\alpha]_D^{20} + 14$  (acetone; c0.33). IR  $\nu_{\text{max}}^{\text{KBr}}$   $\text{cm}^{-1}$ : 3630, 3140, 2995, 2860, 1760, 1740, 1505, 1420, 1380, 1350, 1240, 1180, 1145, 1030, 970, 920, 875, 850, 805, 655, 605.  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR (see Tables 1 and 2). EIMS (direct inlet) 75 eV;  $m/z$  (rel. int.): 404 [M]<sup>+</sup> (18), 312 (38), 369 (22), 220 (28), 218 (28), 187 (30), 159 (21), 157 (19), 96 (98), 95 (88), 94 (20), 91 (40), 81 (48), 43 (100). (Found: C, 65.77; H, 7.12%.  $\text{C}_{22}\text{H}_{28}\text{O}_7$  requires: C, 65.41; H, 7.01%).

*Acetylation of **1** to give **3**.* A soln of 25 mg of 12-epiteupolin II (**1**) in 0.5 ml of pyridine and 0.2 ml of  $\text{Ac}_2\text{O}$  was allowed to stand overnight at room temp. Usual work-up and recrystallization from  $\text{EtOAc}$ - $\text{Et}_2\text{O}$  gave 22 mg of **3**, identical in all respects (mmp,  $[\alpha]_D$ , IR,  $^1\text{H}$  NMR,  $^{13}\text{C}$  NMR, MS, TLC) with natural montanin C.

Table 2.  $^{13}\text{C}$  NMR chemical shifts of compounds **1** and **2** ( $\text{CDCl}_3$ , TMS as int. standard).

C	<b>1</b>	<b>2</b>	$\Delta\delta$
1	22.1 <i>t</i>	22.8 <i>t</i>	—0.7
2	24.9 <i>t</i>	24.9 <i>t</i>	0.0
3	31.9 <i>t</i> * <sup>†</sup>	31.8 <i>t</i> * <sup>†</sup>	+0.1
4	65.5 <i>s</i>	65.2 <i>s</i>	+0.3
5	46.2 <i>s</i>	46.3 <i>s</i>	—0.1
6	73.9 <i>d</i>	73.7 <i>d</i>	+0.2
7	32.7 <i>t</i> * <sup>†</sup>	32.4 <i>t</i> * <sup>†</sup>	+0.3
8	40.7 <i>d</i>	37.9 <i>d</i>	+2.8
9	51.2 <i>s</i>	50.8 <i>s</i>	+0.4
10	50.7 <i>d</i>	52.8 <i>d</i>	—2.1
11	43.6 <i>t</i>	71.5 <i>d</i>	+0.3
12	71.4 <i>d</i>	125.1 <i>s</i>	+0.2
13	107.9 <i>d</i>	108.1 <i>d</i>	+0.2
14	144.2 <i>d</i>	144.2 <i>d</i>	0.0
15	139.1 <i>d</i>	139.6 <i>d</i>	—0.5
16	16.8 <i>q</i>	16.5 <i>q</i>	+0.3
17	47.4 <i>t</i>	47.3 <i>t</i>	+0.1
18	61.5 <i>t</i>	60.8 <i>t</i>	+0.7
19	176.1 <i>s</i>	176.3 <i>s</i>	—0.2
OAc	169.6 <i>s</i>	169.5 <i>s</i>	+0.1
	21.3 <i>q</i>	21.2 <i>q</i>	+0.1

\*Assignments bearing the same sign may be reversed.

## REFERENCES

1. Malakov, P. Y., Papanov, G. Y. and Mollov, N. M. (1978) *Tetrahedron Letters* 2025.
2. Malakov, P. Y., Papanov, G. Y. and Mollov, N. M. (1972) *Z. Naturforsch. B*, **34**, 1570.
3. Papanov, G. Y. and Malakov, P. Y. (1981) *Z. Naturforsch. B*, **36**, 112.
4. Malakov, P. Y., Papanov, G. Y. and Ziesche, J. (1982) *Phytochemistry* **21**, 2597.
5. Gacs-Baitz, E., Kajtar, M., Papanov, G. Y. and Malakov, P. Y. (1982) *Heterocycles*, **19**, 539.
6. Malakov, P. Y. and Papanov, G. Y. (1983) *Phytochemistry* **22**, 2791.
7. Papanov, G. Y. and Malakov, P. Y. (1985) *Phytochemistry* **24**, 297.
8. Malakov, P. Y., Papanov, G. Y., Mollov, N. M. and Spassov, S. L. (1978) *Z. Naturforsch. B*, **33**, 789.
9. Malakov, P. Y. and Papanov, G. Y. (1985) *Phytochemistry* **24**, 301.
10. Papanov, G. Y., Malakov, P. Y. and Bohlmann, F. (1981) *Phytochemistry*, **20**, 170.
11. Savona, G., Paternostro, M., Piozzi, F., Hanson, J. R., Hicheck, P. B. and Thomas, S. A. (1979) *J. Chem. Soc. Perkin Trans. I*, 1915.
12. Savona, G., Paternostro, M., Piozzi, F. and Rodriguez, B. (1979) *Tetrahedron Letters* 379.
13. Fayos, J., Fernandez-Gadea, F., Pascual, C., Perales, A., Piozzi, F., Rico, M., Rodriguez, B. and Savona, G. (1984) *J. Org. Chem.*, **49**, 1789.
14. Rodriguez, M., Barluenga, J., Savona, G., Piozzi, F., Servettaz, O. and Rodriguez, B. (1984) *Phytochemistry* **23**,

1465.

15. Pascual, C., Fernandez, P., Garcia-Alvarez, M. C., Hueso-Rodriguez, J. A., Rodriguez, B., Bruno, M., Paternostro, M., Piozzi, F. and Savona, G. (1986) *Phytochemistry* **25**, 715.
16. Gács-Baitz, E., Papanov, G. Y., Malakov, P. Y. and Szilágyi, L. (1987) *Phytochemistry* (in press).
17. Malakov, P. Y., Papanov, G. Y., Mollov, N. M. and Spassov, S. L. (1978) *Trav. Sci. Univ. Plovdiv (Chimie)* **16**, 215.

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## TRITERPENOID FROM *AGRIMONIA PILOSA*

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**Key Word Index**—*Agrimonia pilosa*; Rosaceae; triterpenoid; 19 $\alpha$ -hydroxyursolic acid.

**Abstract**—Two new triterpenoids have been isolated as a methylester from the whole plant of *Agrimonia pilosa*, along with 2 $\alpha$ ,19 $\alpha$ -dihydroxyursolic acid (28-1)  $\beta$ -D-glucopyranoside from the roots of this plant. On the basis of chemical and spectral evidence, the structures were established as 1 $\beta$ ,2 $\alpha$ ,3 $\beta$ ,19 $\alpha$ -tetrahydroxyurs-12-en-28-oic acid and 1 $\beta$ ,2 $\beta$ ,3 $\beta$ ,19 $\alpha$ -tetrahydroxyurs-12-en-28-oic acid.

### INTRODUCTION

*Agrimonia pilosa* Ledeb is widely distributed in Asia. The chemical components of this plant have been extensively examined and agrimonolide [1], luteolin 7-O- $\beta$ -D-glucoside, apigenin 7-O- $\beta$ -D-glucoside [2, 3], agrimol A, B and D [4], agrimophol [5] and tannins [6, 7] were obtained. The antitumour activity of the extracts from the roots of this plant was also reported [8]. We report now on the constituents of *Agrimonia pilosa*.

### RESULTS AND DISCUSSION

The methanol extract of the aerial parts of *A. pilosa* was fractioned by the usual procedure (Experimental) to afford a triterpenoid fraction. This was treated with diazomethane in methanol because of the difficulty of separation, and compounds **1** (45 mg) and **2** (112 mg) were isolated. The  $^1\text{H}$  NMR spectrum of compound **1**  $\text{C}_{31}\text{H}_{50}\text{O}_6$  (EIMS, *m/z* 518) showed the characteristic broad singlet at  $\delta$  2.57, together with the tertiary methyl [ $\delta$  0.68, 0.83, 1.02 (2Me), 1.22 (2Me)], the secondary methyl (0.94, *d*, *J* = 6.4 Hz), the ester methyl (3.60) and the olefinic (5.35, *t*, *J* = 3.4 Hz) protons, all of which suggested a 19 $\alpha$ -hydroxyurs-12-en type of triterpenoid. The olefinic carbon signals ( $\delta$  130.0, C-12; 137.3, C-13) in the  $^{13}\text{C}$  NMR spectrum of **1** also indicated that **1** had an urs-12-en skeleton [9]. Although the hydroxy methine protons were not obvious in the  $^1\text{H}$  NMR spectrum of **1**, an

acetate (**1a**) of **1** showed the signals of three acetoxy and three acetoxy methine groups. The latter of which exhibited two doublets ( $\delta$  4.79, *J* = 10.6 Hz; 4.88, *J* = 9.3 Hz) and a double doublet ( $\delta$  5.22, *J* = 10.6 and 9.3 Hz), and were assignable to C-1 (or 3), C-3 (or 1) and C-2, respectively. Three hydroxy methine carbons were also indicated in the  $^{13}\text{C}$  NMR spectrum of **1** ( $\delta$  74.6, 74.9, 79.9). As the *J*-values of these signals indicated trans-diaxial correlated protons, the three acetoxy must be equatorial. It was concluded that compound **1** was 1 $\beta$ ,2 $\alpha$ ,3 $\beta$ ,19 $\alpha$ -tetrahydroxyurs-12-en-28-oate, and that the natural compound should be originally 1 $\beta$ ,2 $\alpha$ ,3 $\beta$ ,19 $\alpha$ -tetrahydroxyurs-12-en-28-oic acid (**1b**).

The  $^1\text{H}$  NMR spectrum of compound **2** ( $\text{C}_{31}\text{H}_{50}\text{O}_6$ ; EIMS, *m/z* 518) showed a broad singlet at  $\delta$  2.59 similar to that of **1**. As the other signals of the  $^1\text{H}$  NMR and the  $^{13}\text{C}$  NMR spectra of **2** were quite similar to those of **1**, compound **2** was considered to possess the same skeleton as **1**. An acetate (**2a**) of **2** showed in the  $^1\text{H}$  NMR spectrum the signals of three acetoxy and three acetoxy methine protons. These methine protons appeared at  $\delta$  4.70 (*d*, *J* = 3.7 Hz, C-1 or 3), 4.75 (*d*, *J* = 3.7 Hz, C-3 or 1) and 5.44 (*t*, *J* = 3.7 Hz, C-2) compared to those of **1**, and these coupling constants were accommodated on an axial-equatorial-axial correlation for these protons. These findings indicated **2** is 1 $\beta$ ,2 $\beta$ ,3 $\beta$ ,19 $\alpha$ -tetrahydroxyurs-12-en-28-oate, and that the naturally occurring compound should be 1 $\beta$ ,2 $\beta$ ,3 $\beta$ ,19 $\alpha$ -tetrahydroxyurs-12-en-28-oic acid (**2b**).

An acetonide reaction of **2** with acetone and *p*-toluene sulphonic acid yielded two compounds **2d** and **2e** (trace), although the same reaction for **1** gave only the starting

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